

Effect of K⁺-channel Blockers on the Muscarinic- and A₁-adenosine-Receptor Coupled Regulation of Electrically Evoked Acetylcholine Release in the Rat Hippocampus

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It was attempted to clarify the participation of K⁺-channels in the post-receptor mechanisms of the muscarinic and A₁-adenosine receptor-mediated control of acetylcholine (ACh) release in the present study. Slices from the rat hippocampus were equilibrated with [³H]choline and the release of the labelled products was evoked by electrical stimulation (3 Hz, 5 V/cm, 2 ms, rectangular pulses), and the influence of various agents on the evoked tritium-outflow was investigated. Oxotremorine (Oxo, 0.1~10 μM), a muscarinic agonist, and N⁶-cyclopentyladenosine (CPA, 1~30 μM), a specific A₁-adenosine agonist, decreased the ACh release in a dose-dependent manner, without affecting the basal rate of release. 4-aminopyridine (4AP), a specific A-type K⁺-channel blocker (1~100 μM), increased the evoked ACh release in a dose-related fashion, and the basal rate of release is increased by 3 and 100 μM. Tetraethylammonium (TEA), a non-specific K⁺-channel blocker (0.1~10 mM), increased the evoked ACh release in a dose-dependent manner without affecting the basal release. The effects of Oxo and CPA were not affected by 3 μM 4AP co-treatment, but 10 mM TEA significantly inhibited the effects of Oxo and CPA. 4AP (10 μM)- and TEA (10 mM)-induced increments of evoked ACh release were completely abolished in Ca²⁺-free medium, but these were recovered in low Ca²⁺ medium. And the effects of K⁺-channel blockers in low Ca²⁺ medium were inhibited by Mg²⁺ (4 mM) and abolished by 0.3 μM tetrodotoxin (TTX). These results suggest that the changes in TEA-sensitive potassium channel permeability and the consequent limitation of Ca²⁺ influx are partly involved in the presynaptic modulation of the evoked ACh-release by muscarinic and A₁-adenosine receptors of the rat hippocampus.

Key Words: Tetraethylammonium, 4-aminopyridine, Muscarinic Receptor, A₁-adenosine Receptor, Acetylcholine, Hippocampus

INTRODUCTION

It is well known that ACh release is inhibited by stimulating the presynaptic muscarinic (Szerb & Somogyi, 1973; Kilbinger & Wessler, 1980; Briggs & Cooper, 1982) and A₁-adenosine receptors (Vizi, 1984; Choi et al, 1992), but the underlying me-

chanisms of such presynaptic control of transmitter release are incompletely understood (Starke, 1987; Fredholm & Dunwiddie, 1988; Choi & Oh, 1994).

On the other hand, there are reports suggesting that the K⁺- and Ca²⁺-channels are involved in the synaptic transmission elicited by muscarinic (Fredholm, 1990; Clos et al, 1994) and A₁-adenosine (Proctor & Dunwiddie, 1983; Dunwiddie, 1985; Trussell & Jackson, 1985; Madison et al, 1986; Dolphine et al, 1986) receptors activation in the central nervous system (CNS). However, the involvement of ionic

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conductance system in controlling ACh release by the presynaptic muscarinic and A₁-adenosine receptors still remains unclear.

Recently, K⁺-channels may exist in as many as twenty or more separate types in the CNS, and their pharmacologic and therapeutic significance of which are elucidated (Rudy, 1988; Robertson & Steinberg, 1990). And, numerous studies were undertaken to clarify the influence of K⁺-channel modulators upon the release of neurotransmitters in the CNS. The evoked and basal releases of ACh from the hippocampus were increased by 4AP (Drukarch et al, 1989; Fredholm, 1990) and, TEA (Drukarch et al, 1989), and the presynaptic effects of α₂-, muscarinic- and A₁-adenosine receptors stimulation are reduced by this drug (Okada & Ozawa, 1980; Schoffelmer & Mulder, 1983; Fredholm, 1990). Moreover, Zoltay & Cooper (1990) proposed that the rat cortical muscarinic autoreceptor activation lead to opening of presynaptic K⁺-channel, Fredholm (1990) observed that 4AP blocked the ability of carbachol to decrease the evoked release of ACh from hippocampal slices, and Töröcsik & Vizi (1990) found that 4AP prevents the Oxo-induced decrease in evoked release of ACh from cortical slices. On the other hand, however, there are conflicting reports that the hippocampal presynaptic A₁- and muscarinic receptors do not primarily regulate 4AP-dependent K⁺-channels (Fredholm, 1990; Hu & Fredholm, 1991; Dolezal & Wecker, 1991).

The present study, therefore, was designed to delineate the role of K⁺-channel on the evoked ACh release in the rat hippocampus, and to define the involvement of K⁺-channels in post-receptor mechanisms of muscarinic and A₁-adenosine receptors.

METHODS

Slices of 2.5~3.0 mg, 400 μm in thickness, were prepared from the hippocampus of Sprague-Dawley rats of either sex weighing 250~300 gm with a Balzers[®] tissue chopper (Balzers Union Aktiengesellschaft, England) and were incubated in 2 ml of modified Krebs-Henseleit medium containing 0.1 μmol/L [³H]choline for 30 min at 37°C. Subsequently, the [³H]choline-pretreated slices were superfused with medium containing hemicholinium-3 (10 μM) for 140 min at a rate of 0.5 ml/min. The composition (mM) of superfusion medium was 118 NaCl, 4.8 KCl, 1.3

CaCl₂, 1.2 KH₂PO₄, 1.2 MgSO₄, 25 NaHCO₃, 0.57 ascorbic acid, 0.03 Na₂EDTA, and 11 glucose, and the superfusate was continuously aerated with 95% O₂ + 5% CO₂, the pH adjusted to 7.4.

Collection of 5 min fractions (2.5 ml) of the superfusate began 50 min after the superfusion. Electrical stimulations (3 Hz, 5 V/cm, 2 ms, rectangular pulses) for 2 minutes were performed at 60 min (S₁) and 120 min (S₂). Drugs were added between S₁ and S₂ to the superfusion medium. At the end of superfusion, the slices were solubilized in 0.5 ml tissue solubilizer (0.5 N quaternary ammonium hydroxide in toluene). The radioactivity in the superfusates and solubilized tissues was determined by liquid scintillation counter (Beckman LS 5000TD). The fractional rate of tritium-outflow was calculated as tritium-outflow per 5 min divided by the total tritium content in the slice at the start of the respective 5-min period (Hertting et al, 1980). Drug effects on the evoked tritium-outflow were evaluated by calculating the ratio of the outflow evoked by S₂ and by S₁ (S₂/S₁). The influence of drugs on the basal outflow are expressed as the ratio b₂/b₁ between fractional rates of outflow immediately before S₂ (115~120 min) and S₁ (55~60 min).

The following chemicals were used: [³H]choline (72-78 Ci/mmol, Amersham), oxotremorine sesquifumarate (RBI), N⁶-cyclopentyladenosine (RBI) tetraethylammonium bromide (Sigma), 4-aminopyridine (Nakarai), atropine sulfate (Sigma), hemicholinium-3 (Sigma) and tetrodotoxin (RBI). Drugs were dissolved in the medium except tetrodotoxin which was initially dissolved in DMSO and then diluted with the medium.

All results are given as mean ± SEM. Significance of difference between the groups was determined by ANOVA and subsequently by Duncan test (Snedecor and Cochran, 1980).

RESULTS

Effect of Oxo and CPA on [³H]ACh release evoked by electrical stimulation

Hippocampal slices prelabelled with [³H]choline, a [³H]ACh precursor, were superfused with the medium containing 10 μM hemicholinium-3, a choline uptake inhibitor. And in order to eliminate the inhibition of ACh release by activating muscarinic auto receptors,

Table 1. Effect of oxotremorine (Oxo) on the electrically-evoked and basal tritium-outflows from the rat hippocampal slices preincubated with [³H]choline

Oxo before S ₂ (μM)	n	S ₂ /S ₁	b ₂ /b ₁
none	18	0.888 ± 0.062	0.632 ± 0.023
0.1	7	0.736 ± 0.042	0.623 ± 0.093
0.3	10	0.502 ± 0.041***	0.680 ± 0.019
1.0	7	0.230 ± 0.038***	0.625 ± 0.034
3.0	4	0.135 ± 0.046***	0.662 ± 0.058
10.0	4	0.097 ± 0.038***	0.735 ± 0.018

After preincubation, the slices were superfused with medium containing 10 μM hemicholinium-3 and then stimulated twice (S₁, S₂). Oxo was presented 15 min before S₂. Drug effects on basal outflow are expressed as b₂/b₁ between fractional rates of outflows immediately before S₂ (115~120 min) and before S₁ (55~60 min). Mean ± SEM from the number (n) of observation are given. Significant differences from the drug-free control (none) are marked with asterisks (***) (p < 0.001).

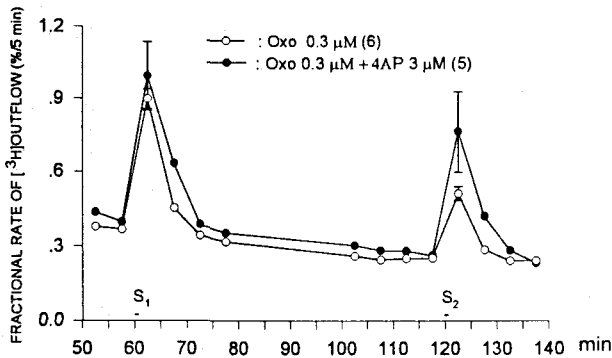


Fig. 1. A typical presentation of the tritium-outflow from the rat hippocampal slices preincubated with [³H]choline. The slices were electrically stimulated twice for 2 min each, after 60 and 120 min of superfusion (S₁, S₂). The drug effect on the stimulation-evoked tritium outflow is expressed by the ratio S₂/S₁ (○; 0.479 ± 0.060 and ●; 0.890 ± 0.128). The radioactivities of the tissues at the start of experiment were 0.656 ± 0.037 (○) and 0.657 ± 0.023 (●) pmol. Oxotremorine (Oxo) and 4-aminopyridine (4AP) were added 15 min before S₂. In the parentheses are the number of experiments.

atropine (30 nM), a muscarinic antagonist, was added in the superfusion medium in the CPA experiments. During superfusion, the tissue was electrically stimulated twice.

As shown in Fig. 1, 0.3 μM Oxo decreased the electrically-evoked outflow of tritium without any change of the basal release. Oxo (0.1~10 μM) and CPA (1~30 μM) decreased the electrically-evoked ACh release in a concentration-dependent manner

(Table 1 and 2).

Effect of K⁺-channel blockers on ACh release

As shown in Table 3, 4AP, a specific A-type K⁺-channel blocker, in doses ranging from 1 to 100 μM, increased the evoked ACh release in a dose-related fashion. Moreover, basal outflow of ACh is significantly increased by 3 and 100 μM 4AP. TEA, a nonspecific K⁺-channel blocker, in doses ranging from 0.03 to 10 mM, increased the evoked ACh release in a dose-dependent manner, but did not change the basal release (Table 4).

Interaction of K⁺-channel blockers and Oxo & CPA on ACh release

To ascertain whether the Oxo and CPA effects are mediated by K⁺-channel modulation, the effects of K⁺-channel blockers upon the Oxo and CPA were then studied.

As shown in Figs. 1 and 2A, when the slices were treated with combination of Oxo and 4AP, 4AP did not affect the inhibitory effects of Oxo effect, except for a slight inhibition of the effect of 1 μM Oxo. And also, the inhibitory effects of CPA were not affected by 4AP co-treatment (Fig. 3A).

Next, the influence of TEA on the Oxo and CPA effects were investigated. As depicted in Figs. 2B and 3B, the Oxo and CPA effects are significantly inhibited by 3 mM TEA.

Table 2. Effect of N⁶-cyclopentyladenosine (CPA) on the electrically-evoked and basal tritium-outflows from the rat hippocampal slices preincubated with [³H]choline

CPA before S ₂ (μM)	n	S ₂ /S ₁	b ₂ /b ₁
none	7	0.975 ± 0.026	0.682 ± 0.025
1	5	0.682 ± 0.096*	0.670 ± 0.026
3	5	0.607 ± 0.039***	0.690 ± 0.013
10	5	0.434 ± 0.042***	0.680 ± 0.025
30	5	0.326 ± 0.058***	0.645 ± 0.019

Significant differences from the drug-free control are marked with asterisks (*p < 0.05). Other legends are the same as in Table 1.

Table 3. Effect of 4-aminopyridine (4AP) on the electrically-evoked and basal outflows of tritium from the rat hippocampal slices preincubated with [³H]choline

4AP before S ₂ (μM)	n	S ₂ /S ₁	b ₂ /b ₁
none	4	0.953 ± 0.064	0.673 ± 0.020
1	5	1.114 ± 0.035	0.749 ± 0.053
3	5	1.349 ± 0.067**	0.763 ± 0.015**
10	5	1.612 ± 0.071***	0.712 ± 0.034
30	5	2.269 ± 0.205***	0.758 ± 0.036
100	5	3.316 ± 0.433***	0.960 ± 0.019***

Significant differences from the drug-free control are marked with asterisks (**p < 0.01). Other legends are the same as in Table 1.

Table 4. Effect of tetraethylammonium (TEA) on the electrically-evoked and basal outflows of tritium from the rat hippocampal slices preincubated with [³H]choline

TEA before S ₂ (mM)	n	S ₂ /S ₁	b ₂ /b ₁
none	4	0.811 ± 0.065	0.696 ± 0.059
0.03	5	1.007 ± 0.029*	0.714 ± 0.026
0.10	5	0.998 ± 0.031*	0.697 ± 0.022
1	5	1.317 ± 0.056**	0.642 ± 0.027*
3	5	1.599 ± 0.049	0.712 ± 0.035
10	5	2.052 ± 0.478	0.716 ± 0.021

Legends are the same as in Table 2.

To clarify the mechanism of the K⁺-channel blockers on the evoked ACh release, the effects of K⁺-channel blockers were investigated in another sets of experiments in which either the concentration of Ca²⁺ was lowered 1.3 to 0.325 or 0 mM, and Mg²⁺ increased from 1.2 to 4 mM. In the preliminary

experiments, the electrically-evoked ACh release was significantly decreased by lowered external Ca²⁺ from 1.3 to 0.325 mM (S₂/S₁; 0.120 ± 0.008, n=4) compared with the control (S₂/S₁; 0.996 ± 0.078, n=8). Adding magnesium (4 mM) to the lowered Ca²⁺ medium 45 min before S₂ onwards, inhibited further

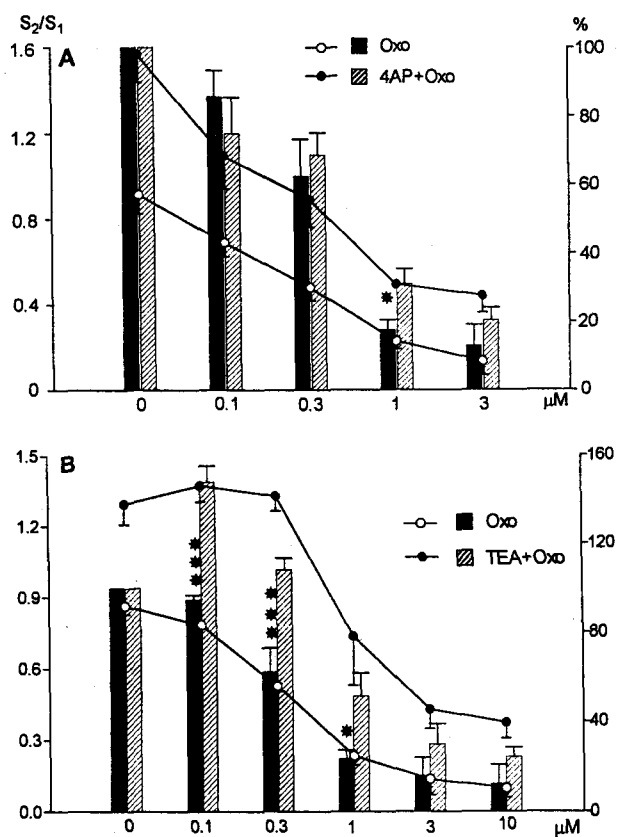


Fig. 2. Influence of the 4-aminopyridine (4AP) 3 μM (A) and tetraethylammonium (TEA) 3 mM (B) upon the effect of oxotremorine (Oxo) on the electrically evoked ACh release from the rat hippocampal slices. Line plots indicate S₂/S₁ ratios. Each point represents mean value from 5-7 experiments, with one SEM as bar. Histograms show percentages of the corresponding controls. Statistically significant difference is indicated with asterisks (*p < 0.05 and ***p < 0.001).

the evoked ACh (S₂/S₁; 0.024 ± 0.009). As shown in Fig. 4, 4AP-induced increments of the evoked ACh release was completely abolished in Ca²⁺-free medium, and this inhibition was lessened in low Ca²⁺ medium with increased basal rate of release. And, increasing effects of evoked and basal rate of release by 4AP in low Ca²⁺ medium were significantly inhibited by Mg²⁺ or completely abolished by TTX added to the medium.

Next, the effects of TEA on the evoked ACh release were investigated in the Ca²⁺ and Mg²⁺ modified media. The TEA-induced increase in evoked ACh release was completely abolished in Ca²⁺-free medium, but significantly recovered in low Ca²⁺ medium. And, this inhibitory effect was also attenuated

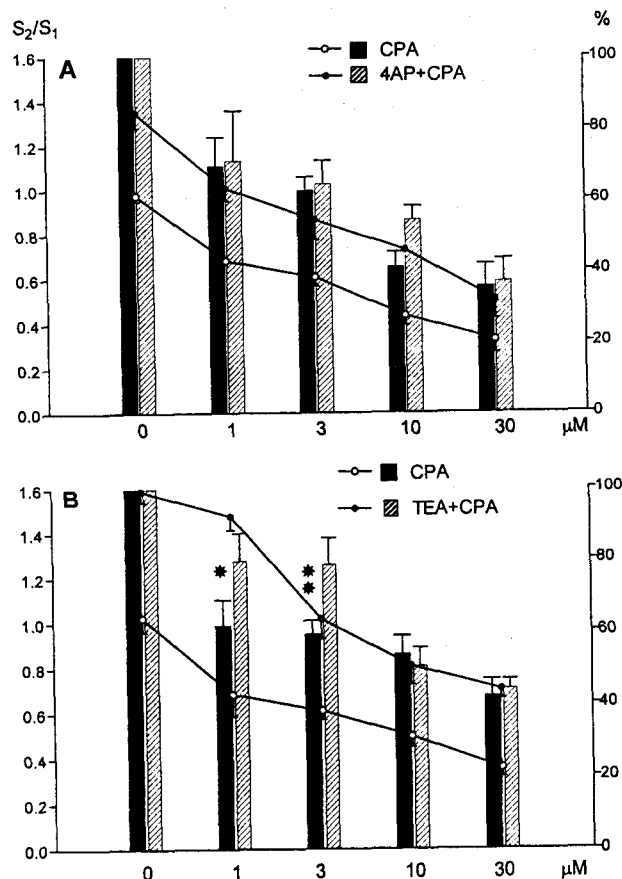


Fig. 3. Influence of the 4-aminopyridine (4AP) 3 μM (A) and tetraethylammonium (TEA) 3 mM (B) upon the effect of N⁶-cyclopentyladenosine (CPA) on the electrically evoked ACh release from the rat hippocampal slices. Statistically significant difference is indicated with *p < 0.01. Other legends are the same as in preceding figures.

by adding Mg²⁺ or abolished by TTX in medium, likewise in 4AP experiments (Fig. 5).

DISCUSSION

A major aim of the present study was to investigate the effects of K⁺-channel blockers to gain more information about the mechanism underlying the pre-synaptic inhibitory effect of muscarinic and A₁-adenosine receptors. In the present study, the electrically evoked release of ACh from the rat hippocampal slice was inhibited by Oxo and CPA. These results are in accordance with other reports that muscarinic agonists such as carbachol & Oxo (Allgaier et al, 1988, 1993a; Choi et al, 1991), and adenosine agonists such as R-N⁶-(2-phenylisopropyl)adenosine, N⁶-cyclopentyl-

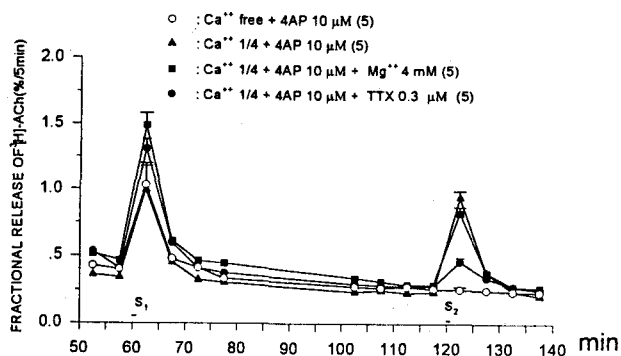


Fig. 4. The influence of Mg^{2+} and tetrodotoxin (TTX) upon the 4AP-induced tritium outflow in lowered or zero external Ca^{2+} concentration from rat hippocampal slices prelabelled with [3H]choline. The concentration of external Ca^{2+} was lowered from 1.3 to 0.325 (1/4) or 0 (Ca^{2+} -free) mmol/l from 15 min and Mg^{2+} was increased 1.2 to 4 mM from 45 min before S_2 onwards, respectively. In some experiments either 4AP or TTX was present from 15 min before S_2 onwards. The S_2/S_1 ratios were: 0.015 ± 0.026 in Ca^{2+} -free; 1.098 ± 0.024 in low Ca^{2+} ; 0.552 ± 0.022 in low Ca^{2+} /high Mg^{2+} ; 0.251 ± 0.034 in low Ca^{2+} /TTX. In the parentheses are the number of experiments.

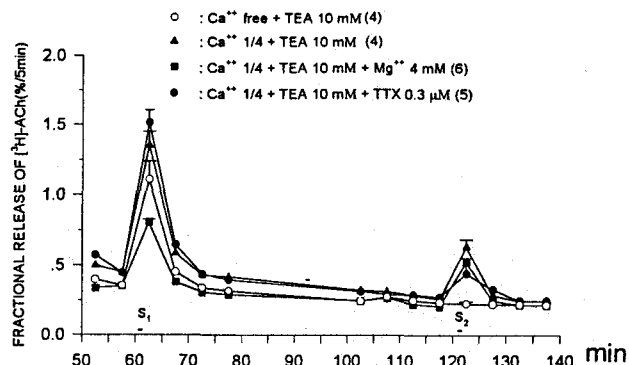


Fig. 5. The influence of Mg^{2+} and TTX upon the TEA-induced tritium outflow in lowered or zero external Ca^{2+} concentration from rat hippocampal slices prelabelled with [3H]choline. The concentration of external Ca^{2+} was lowered from 1.3 to 0.325 (1/4) or 0 (Ca^{2+} -free) mM from 15 min and Mg^{2+} was increased 1.2 to 4 mM from 45 min before S_2 onwards, respectively. In some experiments either TEA or TTX was present from 15 min before S_2 onwards. The S_2/S_1 ratios were: -0.005 ± 0.014 in Ca^{2+} -free; 0.682 ± 0.012 in low Ca^{2+} ; 0.390 ± 0.043 in low Ca^{2+} /high Mg^{2+} ; 0.216 ± 0.019 in low Ca^{2+} /TTX. In the parentheses are the number of experiments.

tyladosine & adenosine (Duñer-Engström & Fredholm, 1988; Choi et al, 1992; Choi & Yoon, 1993; Choi & Oh, 1994) decreased the electrically-evoked release of ACh in the hippocampus.

On the other hand, there are reports that various types of voltage-dependent K^+ -channels exist in the CNS (Rudy, 1988; Storm, 1990) and evidence has been presented indicating that K^+ -channels are involved in the releasing process of neurotransmitters in striatum (Drukarch et al, 1989) and hippocampus (Jackisch et al, 1992; Allgaier et al, 1993b). Therefore, in order to confirm the effects of voltage-dependent K^+ -channel blockers on the evoked ACh release were investigated in this study.

4AP and TEA substantially increased ACh release evoked by electrical stimulation. The increasing effect was abolished by removing Ca^{2+} in the medium, but was significantly recovered in low Ca^{2+} medium. And the inhibitory effect was attenuated by Mg^{2+} and abolished by TTX, indicating that it reflects nerve impulse-triggered and calcium-dependent release. These facts, in conjunction with the evidence that K^+ -channel blockers inhibit the outward K^+ -currents, indicate that the increment of the transmitter release is probably due to increase of Ca^{2+} influx into the axon terminal followed by a prolongation of the

duration of action potential. There are, however, several points not to be overlooked in the action of 4AP. In contrast to the effects of TEA, large doses of 4AP induce by itself significant increase of the spontaneous ACh release in normal and even in low Ca^{2+} medium. And also, 4AP significantly increased the evoked ACh release (S_2/S_1 : 0.552 ± 0.022 , $n=5$, $p<0.001$) compared with the drug-free control in low Ca^{2+} /high Mg^{2+} medium (S_2/S_1 : 0.016 ± 0.026 , $n=5$). In view of the findings that the action site of 4AP is intracellular (Rudy, 1988) and that magnesium ion block the ionic channel in rat hippocampal slices (Mody et al, 1987), the possibility that the increments of spontaneous and evoked ACh release by 4AP may result from intracellular Ca^{2+} release from the storage sites could not be ruled out. But, 4AP dependent increments of evoked and spontaneous release were completely abolished by TTX-treatment and by removal of Ca^{2+} in the medium, indicating that these effects are TTX-sensitive and extracellular Ca^{2+} -dependent. On the basis of these findings, in conjunction with other reports that TEA blocks a wide variety of K^+ -channels, whereas 4AP is supposed to possess some selectivity for the A current and delayed rectifier (for review see: Cook, 1988; Rudy, 1988; Castle et al, 1989), it is most likely that the

TEA- and 4AP-induced ACh release is mediated through different K⁺-channels.

It is generally known that the presynaptic potassium conductance is involved in the feedback regulation of ACh release of the hippocampal ACh release (Zoltay & Cooper, 1990). Therefore, in order to confirm the involvement of voltage dependent K⁺-channels in muscarinic- and A₁-adenosine-receptor-mediated decrease of ACh release, the influence of K⁺-channel blockers upon the Oxo and CPA effects were investigated in this study.

In the interaction experiments, the concentration-response relations for Oxo and CPA observed in the presence of 4AP and TEA. The fact that TEA significantly attenuated the inhibitory effects of Oxo and CPA, but 4AP did not affect them. These results were quite in agreement with the reports that the K⁺-channels are coupled to evoked-ACh release in the rat hippocampus (Fredholm, 1990) and cerebral cortex (Törocsik & Vizi, 1990), suggesting that Oxo and CPA binding to their specific receptors induces the opening of TEA-sensitive K⁺-channels and limits the Ca²⁺ influx. However Fredholm (1990) found that the presynaptic effect of carbachol on muscarinic autoreceptors was abolished by 4AP and proposed that the inhibitory effects of carbachol on hippocampal ACh release are coupled to 4AP sensitive K⁺-channels. Furthermore, Törocsik & Vizi (1990) reported that 4AP interrupts the modulation of ACh release mediated by muscarinic and opiate receptors. In addition, there are reports that depressant effects of adenosine is blocked by 4AP in olfactory cortex (Scholfield and Steal, 1988) and hippocampus (Okada & Ozawa, 1980; Schubert et al, 1986). Discrepancies between the present results and the above reports may not be easily reconciled, but one of the possibilities accounting for the difference is that they did not observe the dose-response relationships of agonists and of agonists under K⁺-channel blockers. And also, they clearly indicated that there are differences in the mechanism underlying different receptor effects and in species. Hence, further studies are required to determine the exact mechanism in the ACh release mediated by presynaptic muscarinic and A₁-adenosine receptors in the rat hippocampus.

Taken together, the present study showed that the decrements of the evoked ACh release by muscarinic and A₁-adenosine receptors is related to the TEA-sensitive K⁺-channels, but not to the 4AP-sensitive K⁺-channels, and that the changes of Ca²⁺ influx are

consequential to this.

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