# Importance of The Location of The Negative-charged Counter-ion against The Protonated Schiff Base on The Chromophore Configuration of *pharaonis*Phoborhodopsin

Kazumi Shimono\*, Yukako Ikeura, Yuki Sudo, Masayuki Iwamoto and Naoki Kamo Laboratory of Biophysical Chemistry, Graduate School of Pharmaceutical Sciences, Hokkaido University, Sapporo, 060-0812, Japan

pharaonis phoborhodopsin (ppR), a photophobic sensor of haloalkaliphilic bacteria, Natronobacterium pharaonis, has retinal as a chromophore covalently bound to Lys in G-helix via a protonated Schiff base (PSB), as is the same as bacteriorhodopsin (bR). For ppR, the corresponding counter-ion is Asp residue (Asp75) located in C-helix. Here we investigated the influence of the protonated state of this counter-ion and its location on the chromophore configuration. Under alkaline condition, the chromophore configuration of D75E mutant was analyzed by HPLC. D75E had a much larger content of 13-cis isomer: the ratio of 13-cis to all-trans was 6:4 while the wild-type had this ratio of 1:9. On the other hand, under acidic condition where Glu was associated, D75E had no 13-cis retinal isomer. Mutants whose Asp75 was replaced by neutral amino acids (D75N and D75Q) did not contain 13-cis retinal. Furthermore, retinal isomer compositions and the change in the visible absorption spectra (indicating the dissociation state of Glu75) were measured under varying pH, and these were almost the same dependencies. These results indicate that an important factor determining the 13-cis isomer content is the presence of negative charge of the counter-ion against PSB, but not the size of this residue. Comparison between the wild-type and D75E in alkaline solutions indicates the influence of the location of the counter-ion.

Key words: photosensor, archaeal retinal protein, retinal isomer, counter-ion, protonated Schiff base

## INTRODUCTION

Haloarchaea, Halobacterium salinarum has at least four retinal proteins: bacteriorhodopsin (bR) [1], halorhodopsin (hR) [2], sensoryrhodopsin (sR or sRI) [3] and phoborhodopsin (pR also called sensory rhodopsin II, sRII) [4]. The former two proteins work as light-driven ion pumps. The latter two work as photosensor and control the motion of this bacteria against light. It is covalently bound to a retinal chromophore at a conserved lysine residue on G helix via a protonated Schiff base (PSB) bond. In the dark, light-driven ion pumps (bR and hR) have all-trans and 13-cis retinal, whereas photosensors (sR and pR) only all-trans [5]. This difference of retinal isomer composition may be related to the difference of function (pump or sensor).

pR-like protein (pharaonis phoborhodopsin, ppR) from haloalkaliphilic bacterium, Natronobacterium pharaonis, is stable under low ionic strength condition and in detergent micelle [6]. Furthermore, we succeeded in the functional expression of ppR in the Escherichia coli membrane [7]. These features are suitable on the study of photosensor

proteins. As is previous Fourier transform Infrared (FT-IR) study shown, in M-intermediate of ppR, Asp75 (corresponding to Asp85 of bR) is proton acceptor from PSB (Y. Furutani, unpubilished and ref. 8). Furthermore, D75N mutant has large red shifted absorption spectrum, but D201N spectrum has no alternation from the wild-type (data not shown). Therefore, Asp75 acts as a counter-ion for PSB. Recently, two groups reported the crystallographic structure of ppR [9,10]. The structure around chromophore in ppR is similar to that in bR. However, the hydrogen bond between PSB and counter-ion or internal water molecule in ppR is stronger than that in bR according to lowtemperature FT-IR spectroscopy [11]. We previously constructed a multiple mutant of ppR (BR/ppR) whose all amino acid residues of the retinal binding pocket were replaced simultaneously with those of bR, and reported that the hydrogen bond of PSB in BR/ppR remained strong from FT-IR study [12,13]. These facts may indicate that the environment around PSB in ppR slightly differs from that in bR.

Here we report that influence of the size and electric charged state of the counter-ion on the chromophore configuration of ppR.

E-mail: kaz@pharm.hokudai.ac.jp

<sup>\*</sup>To whom correspondence should be addressed.

#### MATERIALS AND METHODS

Sample preparation. The genes for the single replacements were prepared by QuickChange Site-Directed Mutagenesis Kit (STRATAGENE), and then pET/ppRHis was used as template [11]. The expression of histidine-tagged recombinant ppRs in *E.coli* BL21(DE3) and its purification were described elsewhere [11].

Spectroscopic titrations. For spectroscopic titration in the absence of Cl, the ppR sample was suspended in 6 mix buffer (citric acid, MES, HEPES, MOPS, CHES and CAPS whose concentrations were 10 mM each) and 0.1 % n-dodecyl-β-D-maltoside (DM). The pH was first adjusted to almost 9 with 10 N NaOH. The titration was carried out from alkaline to acidic pH by adding 1 N H<sub>2</sub>SO<sub>4</sub>.

HPLC analysis. A high performance liquid chromatograph (HPLC) was applied as described previously [12]. A sample was suspended in 6 mix buffer and 0.1 % DM adjusted appropriate pH.

### RESULTS AND DISCUSSION

pH dependency of the pigment spectrum.

The spectrum of D75E was greatly pH-dependent because pKa of Glu-75 was raised to 6.2 while that of wild-

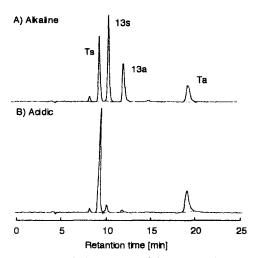


Figure 1. HPLC elution pattern of the chromophores extracted in dark from D75E ppR mutants in the alkaline (A) and acidic condition (B). The detection beam was set at 360 nm. The assignment of each peaks was done by comparison with HPLC pattern of the chromophores extracted from bR. The pH of a sample medium was adjusted 9 (A) or 5 (B). Ts, Ta, 13s and 13a stand for all-trans 15-syn retinal oxime, all-trans 15-anti retinal oxime, 13-cis 15-syn retinal oxime and 13-cis 15-anti retinal oxime, respectively.

type Asp-75 was 3.6 (Fig. 2 and ref. 15). For bR, the replacement of Asp-85 to Glu also raises pKa of the counterion, but the shift is larger than that of ppR [16]. This may indicate the difference of circumstances around the chromophore between bR and ppR; there exists the "pentagon structure" involving PSB, Asp85 and water molecules in the wild-type bR and ppR. In the ppR, this pentagon structure is perturbed relatively to that in bR [9,10,17]. Replacement with Glu might weaken this structure, which raises pKa. Therefore, Glu-replacement of ppR raises pKa by relatively small value.

Retinal configuration of Asp75 mutant.

Figure 1 shows HPLC elution pattern of the chromophore extracted in the dark from D75E mutant. Glu75 is almost dissociated at pH 9 (Fig. 2). At this condition, D75E mutant had a much larger content of 13-cis isomer than the wild-type: ratio of 13-cis to all-trans retinal is 6:4 for D75E, 1:9 for wild-type (Fig. 1A). Next, we investigated whether this existence of the stable 13-cis isomer is originated from the size or the charge of the introduced glutamate residue. In acidic condition where Glu is protonated, the 13-cis content decreases (Fig. 1B). Furthermore, we analyzed the retinal configuration of mutants whose Asp-75 was replaced by neutral amino acids (D75N and D75Q). D75N and D75Q does not have 13-cis isomer (data not shown). These observations suggest that the charged state of the counterion (Glu75) determines the composition of the retinal isomer; under the condition of Glu75 being deprotonated, the

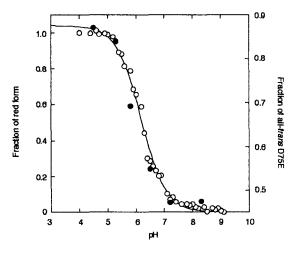


Figure 2. pH dependencies of absorption spectrum (open circle) and the ratio of all-trans isomer determined by HPLC (closed circle) in D75E. For determining the fraction of red form, the increase in absorbance at 542 nm was plotted as the fraction of its maximum value for the preparation as a function of pH. The curves correspond to the equation for a monoprotic titration with pKa values determined by a fit to the data.

pigment has 13-cis retinal while the 13-cis content decreases when the position of Glu75 become neutralized.

To confirm this hypothesis, we investigated the pH dependency of the retinal isomer composition of D75E mutant under varying pH. The pH dependency of retinal isomer composition was almost the same to that of the absorption spectrum which indicates the degree of protonated state of Glu75 (Fig. 2). This indicates that the negative charge of counter-ion is necessary for the existence of stable 13-cis chromophore. The distance between PSB and the counterion depends on the retinal isomer; for 13-cis isomer, the distance is shorter than that for all-trans isomer. When the counter-ion is deprotonated, the larger electrical interaction between this negative charge and the positive PSB stabilizes more 13-cis conformation than all-trans isomer. Therefore, when the counter-ion is protonated, PSB is not stable, and the 13-cis form is not stable. This is consistent that in the case of bR, the all-trans isomer composition is increased under acidic condition [18].

Since the wild-type ppR has all-trans retinal with Asp75, the counter-ion being deprotonated, this rule seems not to hold for ppR. However, careful examination shows that the 13-cis content seems to increase where Asp75 is deprotonated (data not shown). The reason why the effect of the charged state is small in the wild-type might be the longer distance between PSB and the counter-ion of the wild-type than D75E by one methylene chain.

# REFERENCE

- 1. Lanyi, J. K. and H. Luecke (2001). Bacteriorhodopsin. Curr. Opin. Struct. Biol. 11, 415-419.
- 2. Varo, G. (2000). Analogies between halorhodopsin and bacteriorhodopsin. *Biochim. Biophys. Acta* 1460, 220-229.
- 3. Hoff, W. D., K. H. Jung and J. L. Spudich (1997). Molecular mechanism of photosignaling by archaeal sensory rhodopsins. *Annu. Rev. Biophys. Biomol. Struct.* 26, 223-258.
- Sasaki, J. and J. L. Spudich (2000). Proton transport by sensory rhodopsins and its modulation by transducerbinding. *Biochim. Biophys. Acta* 1460, 230-239.
- 5. Imamoto, Y., Y. Shichida, J. Hirayama, H. Tomioka, N. Kamo and T. Yoshizawa (1992). Chromophore configuration of *pharaonis* phoborhodopsin and its isomerization on photon absorption. *Biochemistry* 31, 2523-2528.
- 6. Kamo, N., K. Shimono, M. Iwamoto and Y. Sudo (2001). Photochemistry and Photoinduced Proton-Transfer by *Pharaonis* Phoborhodopsin. *Biochemistry* (Mosc) 66,

- 1277-1282.
- 7. Shimono, K., M. Iwamoto, M. Sumi and N. Kamo (1997). Functional expression of *pharaonis* phoborhodopsin in *Escherichia coli*. *FEBS Lett.* 420, 54-56.
- 8. Engelhard, M., B. Scharf and F. Siebert (1996). Protonation changes during the photocycle of sensory rhodopsin II from *Natronobacterium pharaonis*. FEBS Lett. 395, 195-198
- 9. Luecke, H., B. Schobert, J. K. Lanyi, E. N. Spudich and J. L. Spudich (2001). Crystal Structure of Sensory Rhodopsin II at 2.4 A: Insights into Color Tuning and Transducer Interaction. *Science* 293, 1499-1503.
- 10. Royant, A., P. Nollert, K. Edman, R.Neutze, E. M.Landau, E.Pebay-Peyroula and J. Navarro (2001). X-ray structure of sensory rhodopsin II at 2.1-A resolution. *Proc. Natl. Acad. Sci. USA* 98, 10131-10136.
- 11. Kandori, H., K. Shimono, Y. Sudo, M. Iwamoto, Y. Shichida and N. Kamo (2001). Structural changes of *pharaonis* phoborhodopsin upon photoisomerization of the retinal chromophore: Infrared spectral comparison with bacteirorhodopsin. *Biochemistry* 40, 9238-9246.
- 12. Shimono, K., Y. Ikeura, Y. Sudo, M. Iwamoto and N. Kamo (2001). Environment around the chromophore in *pharaonis* phoborhodopsin: mutation analysis of the retinal binding site. *Biochim. Biophys. Acta* 1515, 92-100.
- 13. Shimono, K., Y. Furutani, H. Kandori and N. Kamo (2002). A *pharaonis* Phoborhodopsin Mutant with the Same Retinal Binding Site Residues As in Bacteriorhodopsin. *Biochemistry* 41, 6504-6509.
- 14. Mowery, P. C., R. H. Lozier, Q. Chae, Y. W. Tseng, M. Taylor and W. Stoeckenius (1979). Effect of acid pH on the absorption spectra and photoreactions of bacteriorhodopsin. *Biochemistry* 18, 4100-4107.
- 15. Shimono, K., M. Kitami, M. Iwamoto and N. Kamo (2000). Involvement of two groups in reversal of the bathochromic shift of *pharaonis* phoborhodopsin by chloride at low pH. *Biophys. Chem.* 87, 225-230.
- 16. Lanyi, J. K., J. Tittor, G. Varo, G. Krippahl and D. Oesterhelt (1992). Influence of the size and protonation state of acidic residue 85 on the absorption spectrum and photoreaction of the bacteriorhodopsin chromophore. *Biochim. Biophys. Acta* 1099, 102-110.
- 17. Luecke, H., B. Schobert, H. T. Richter, J. P. Cartailler and J. K. Lanyi (1999). Structure of bacteriorhodopsin at 1.55 A resolution. *J. Mol. Biol.* 291, 899-911.
- 18. Balashov, S. P., E. S. Imasheva, R. Govindjee, M. Sheves and T. G. Ebrey (1996). Evidence that aspartate-85 has a higher pK(a) in all-trans than in 13-cis bacteri-orhodopsin. *Biophys. J.* 71, 1973-1984.