

가

## Pathogenesis of Inflammatory Muscle Diseases

Dong Kuck Lee, M.D.

Department of Neurology, School of Medicine, Catholic University of Daegu

The inflammatory myopathies are divided into three major and distinct subsets as polymyositis(PM), dermatomyositis(DM), and inclusion body myositis(IBM). This distinction is based on unique clinical, demographic, laboratory, histologic, therapeutic, prognostic, and immunopathologic criteria.

Although the causes of PM, DM, and IBM are unknown, autoimmune mechanisms are implicated, as supported by their association with other putative or definite autoimmune diseases or viruses, the evidence for a T cell-mediated myocytotoxicity or complement-mediated microangiopathy, the presence of various autoantibodies and their response to immunotherapies. But in IBM the immune-mediated process is weaker and IBM patients do not readily respond to immunotherapies, there are convincing immunopathological signs to suggest that a definite autoimmune component, similar to that seen in PM, also plays a role in the cause of IBM.

**Key Words :** Polymyositis, Dermatomyositis, Inclusion body myositis

(Table 2)<sup>1-6</sup>

(infiltrates)

가

10 1

가 (Table 1)  
(polymyositis),  
(inclusion body

(dermatomyositis),  
myositis)

가

<sup>1,2</sup>

**Table 1.** The inflammatory myopathies<sup>1</sup>

Idiopathic	
(1)	Dermatomyositis
(2)	Polymyositis
(3)	Inclusion body myositis
Associated with collagen vascular diseases	
(1)	Systemic lupus erythematosus
(2)	Mixed connective tissue disease
(3)	Scleroderma
(4)	Sjögren's syndrome
(5)	Rheumatoid arthritis
Infective	
(1)	Viral
(2)	Parasitic
(3)	Bacterial
(4)	Fungal
Miscellaneous	
(1)	Eosinophilic myositis
(2)	Associated with vasculitis
(3)	Granulomatous
(4)	Graft versus host disease
(5)	Macrophagic myofasciitis

Address for correspondence

**Dong-Kuck Lee, M.D., PhD.**

Department of Neurology, School of Medicine, Catholic University of Daegu, 3056-6 Daemyung 4-dong, Nam-gu, Daegu 705-718, Korea

Tel : +82-53-650-4267 Fax : +82-53-654-9786

E-mail : dklee@cataegu.ac.kr

**Table 2.** Clinical and Laboratory Features Associated with Inflammatory Myopathies<sup>2,6,8</sup>

Characteristic	Polymyositis	Dermatomyositis	Inclusion Body Myositis
Age of onset	>18 yr	Adulthood and childhood	>50 yr
Sex	Female>male	Female>male	Male>female
Development of muscle symptoms	Subacute	Acute	Slowly
Predominant involvement of muscle weakness	Proximal muscles	Proximal muscles	Proximal and distal muscles
Muscle wasting	Present in chronic forms	Not prominent	Nearly always pronounced in selected muscles (triceps, finger flexors, quadriceps)
Myalgia	Sometimes	Often (especially in acute cases)	Never
Rash or calcinosis	Absent	Present	Absent
Familial association	No	No	Yes, in some cases <sup>a</sup>
Creatine kinase	Increased (up to 50 × normal)	Increased (up to 50 × normal)	Normal or mildly increased (<10 × normal)
Muscle biopsy	Endomysial inflammation	Perimysial and perivascular inflammation; membrane attack complex, immunoglobulin, complement deposition on vessels	Endomysial inflammation; rimmed vacuoles; amyloid deposits; electron microscopy: 15- to 18-nm tubulofilaments
Cellular infiltrate	CD8 <sup>+</sup> T cells; macrophages	CD4 <sup>+</sup> T cells; B cells	CD8 <sup>+</sup> T cells; macrophages
Response to immunosuppressive therapy	Yes	Yes	None or minimal
Associated conditions			
Connective tissue diseases	Yes <sup>a</sup>	Scleroderma and mixed connective tissue disease (overlap syndromes)	Yes, in up to 20% of cases
Other autoimmune diseases <sup>b</sup>	Frequent	Infrequent	Rare, but more frequently recognized
Malignancy	No	Yes, in up to 15% of cases	No
Viruses	Yes, with HIV, HTLV-I, <sup>c</sup> other viruses are uncertain	Unproven	Unproven (rare cases with HIV, HTLV-1)
Drugs <sup>d</sup>	Yes	Yes, rarely	No

a Systemic lupus erythematosus, rheumatoid arthritis, Sjögren's syndrome, systemic sclerosis, mixed connective tissue disease

b Crohn's disease, vasculitis, sarcoidosis, primary biliary cirrhosis, adult celiac disease, discoid lupus, ankylosing spondylitis, Behcet's syndrome, myasthenia gravis, acne fulminans, chronic graft-versus-host disease, dermatitis herpetiformis, psoriasis, Hashimoto's disease, granulomatous diseases, agammaglobulinemia, monoclonal gammopathy, hypereosinophilic syndrome, Lyme disease, Kawasaki disease, autoimmune thrombocytopenia, hypergammaglobulinemic purpura, hereditary complement deficiency, IgA deficiency.

c HTLV-I, human T cell lymphotropic virus type I.

d Drugs include penicillamine(polymyositis and dermatomyositis), zidovudine(polymyositis), and contaminated tryptophan(dermatomyositis-like illness). Other myotoxic drugs may cause myopathy but not an inflammatory myopathy.

가

3

50

가

driceps)

(qua-

가

60% 가

가 가

T CD8 가  
CD8 T 가  
MHC-I

(sarcolemma)  
MHC-I (macrophage) T cytokine 가 cytokine  
T cytokine  
(interleukin 2, 4, 5, interferon ),  
cytokine(interleukin 1, 6, TNF ),  
(L-selectin, integrin LFA-1, VLA-4)  
(GlyCAM-1, ICAM-1, VCAM-1)

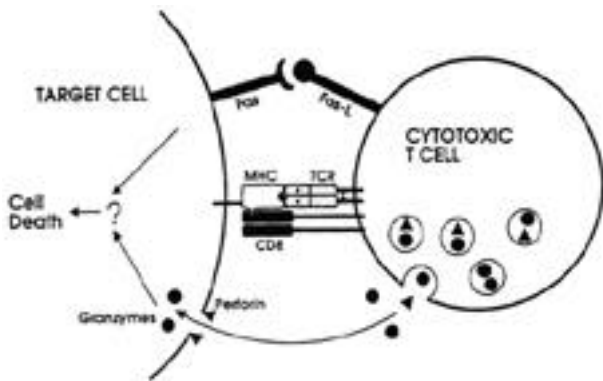
T 가  
(transmigration)  
metalloproteinase(MMP-2, MMP-9) T 가  
T4

T8 T 가 B  
CD8 T 가 (basal lamina)

(invasion)  
perforin granzyme

가 가  
, T 가 가  
가  
가 (self-sensitization)

1. (Fig. 1)<sup>1,4,6-16</sup>



**Figure 1.** Schematic illustration of two major pathways of T-cell-mediated cytotoxicity. One pathway depends on the secretion of perforin- and granzyme-containing cytotoxic granules (black triangles and dots, respectively)(bottom). The other pathway depends on ligation of the Fas death-receptor expressed on the target cell (top). Prior to activation of the cytotoxic pathways, the T-cell receptor (TCR) of the CD8<sup>+</sup> cytotoxic T cell reacts with an antigenic peptide bound to a major histocompatibility complex (MHC) class I molecule on the target cell.<sup>9</sup>

가 가

2. <sup>1,3,4,6,7,10-12</sup>

B 가 B  
CD4<sup>+</sup> 가 CD8<sup>+</sup> 가  
가 CD4<sup>+</sup> 가  
가  
membrane attack complex가  
(humoral)

C5b-9 membranolytic attack complex

가

가



**Table 3.** Autoantibodies in inflammatory myopathies<sup>s</sup>

	Frequency	Conditions
<b>Muscle-specific antibody</b>		
Antisynthetase		
Jo-1	18~20%	Polymyositis and dermatomyositis with interstitial lung disease, arthritis, Raynaud phenomenon, "mechanic's hands," moderate response to therapy
Others (PL-7, PL-12, EJ, OJ)	3%	Same as Jo-1
Nonsynthetase		
Signal recognition particle	4%	Polymyositis with acute onset/severe weakness, poor response to therapy
Mi-2	15~20%	Dermatomyositis with good response to therapy
Mas	1%	Polymyositis with alcoholic rhabdomyolysis; chronic hepatitis
<b>Associated with overlap syndromes</b>		
Antinuclear antibodies	16~32%	All myositis groups
	24~62%	Dermatomyositis
	16~40%	Polymyositis
	72~77%	Overlap syndromes
	19~23%	Inclusion body myositis
Polymyositis-Scl	<8%	All myositis groups
	25%	Scleroderma-myositis (North America)
Anti-Ku	1%	Scleroderma-myositis (Japan)
		Systemic lupus erythematosus without myositis (North America)
U1 ribonuclear protein	12%	Mixed connective tissue disease, systemic lupus erythematosus
SS-A/SS-B	90%	Sjögren syndrome
	10%	Systemic lupus erythematosus

haplotype 가 (allele)가 retrovirus와 HIV-1 zidobudine

5. 4,6-8

coxsackievirus, paramyxovirus, Epstein - Barr 가

sackievirus molecular mimicry cox- 가 in situ

가 hybridization

Jo-1 가 histidyl-transfer RNA synthetase picornavirus genomic RNA

가 (PCR)

retrovirus 가

(HIV) T 가

(HTLV) HIV HTLV -1 MHC-1 Jo -1

가 ?

가 ?

가 ?

가 ?

가 HIV-1 HTLV -1

## REFERENCES

1. Hilton-Jones D. Inflammatory muscle diseases. *Curr Opin Neurol* 2001;14:591-596.
2. Pongratz D, Dalakas MC. Inflammatory myopathies. In: Brant T, Caplan LR, Dichgans J, Christoph Diener H, Kennard C. *Neurological disorders*. 1st ed. New York: Academic Press. 1996;965-969.
3. Bratt R, Shannon KM. Autoimmune and inflammatory disorders. In: Goetz CG, Pappert EJ. *Textbook of clinical neurology*. 1st ed. Philadelphia: W.B. Saunders Company. 1999;1026-1028.
4. Brooke MH. Disorders of skeletal muscle. In: Bradley WG, Daroff RB, Fenichel GM, Marsden CD. *Neurology in clinical practice*. 3rd ed. Vol. 2. Boston: Butterworth Heinemann. 2000;2223-2228.
5. Victor M, Ropper AH. *Principles of neurology*. 7th ed. New York: McGraw-Hill. 2001;1482-1489.
6. Dalakas MC. Polymyositis, dermatomyositis, and inclusion body myositis. In: Brauwald E, Fauci AS, Kasper DL, Hauser SL, Longo DL, Jameson JL. *Principles of internal medicine*. 15th ed. Vol. 2. New York: McGraw-Hill. 2001; 2524-2529.
7. Dalakas MC. Immunopathogenesis of inflammatory myopathies. *Ann Neurol* 1995;37(S1):S74-S86.
8. Amato AA, Barohn RJ. Inflammatory myopathies: dermatomyositis, polymyositis, inclusion body myositis, and related diseases. In: Schapira AH, Griggs RC. *Muscle diseases*. 1st ed. Boston: Butterworth Heinemann. 1999;299-317.
9. Behrens L, Bender A, Johnson MA, Hohlfeld R. Cytotoxic mechanisms in inflammatory myopathies: Co-expression of Fas and protective Bcl-2 in muscle fibers and inflammatory cells. *Brain* 1997;120:929-938.
10. Hohlfeld R, Engel AG, Goebels N, Behrens L. Cellular immune mechanisms in inflammatory myopathies. *Curr Opin Neurol* 1997;9:520-526.
11. Nagaraju K, Raben N, Loeffler L. Conditional up-regulation of MHC class I in skeletal muscle leads to self-sustaining autoimmune myositis and myositis-specific autoantibodies. *Proc Natl Acad Sci USA* 2000;97:9209-9214.
12. Arahata K, Engel AG. Monoclonal antibody analysis of monoclonal cells in myopathies. I: quantitation of subsets according to diagnosis and sites of accumulation and demonstration and counts of muscle fibers invaded by T cells. *Ann Neurol* 1984;16:193-208.
13. Angel AG, Arahata K. Monoclonal antibody analysis of mononuclear cells in myopathies. II: phenotypes of autoinvasive cells in polymyositis and inclusion body myositis. *Ann Neurol* 1984;16:209-215.
14. Arahata K, Engel AG. Monoclonal antibody analysis of mononuclear cells in myopathies. III: immunoelectron microscopy aspects of cell-mediated muscle fiber injury. *Ann Neurol* 1986;19:112-125.
15. Arahata K, Engel AG. Monoclonal antibody analysis of mononuclear cells in myopathies. IV: cell-mediated cytotoxicity and muscle fiber necrosis. *Ann Neurol* 1988;23: 168-173.
16. Arahata K, Engel AG. Monoclonal antibody analysis of mononuclear cells in myopathies. V: identification and quantitation of T8<sup>+</sup> cytotoxic and T8<sup>+</sup> suppressor cells. *Ann Neurol* 1988;23:493-499.
17. Askanas V, Alvarez RV, Mirabella M, Engel WK. Use of anti-neurofilament antibody to identify paired-helical filaments in inclusion body myositis. *Ann Neurol* 1996;39:389-391.
18. Askanas V, Engel WK. Inclusion body myositis: newest concepts of pathogenesis and relation to aging and Alzheimer disease. *J Neuropathol Exp Neurol* 2001;60:1-14.
19. Banwell BL, Engel AG. AlphaB-crystallin immunolocalization yields new insights into inclusion body myositis. *Neurology* 2000;54:1033-1041.
20. Moslemi AR, Lindberg C, Oldfors A. Analysis of multiple mitochondrial DNA deletions in inclusion body myositis. *Hum Mutat* 1997;10:381-386.
21. Mozaffar T, Pestronk A. Myopathy with anti-Jo-1 antibodies: pathology in perimysium and neighbouring muscle fibres. *J Neurol Neurosurg Psychiatry* 2000;68:472-478.