

Pethidine induced changes in ovarian follicular kinetics and biochemical parameters in albino rats

Vijaykumar B Malashetty^{1,*}, Somanath Reddy Patil² and Saraswati B Patil²

¹*Department of Biology, Sharnbasveshwar Independent P.U. College of Science, Gulbarga-585 102, India;*

²*Laboratory of Reproductive Biology, Department of Zoology, Gulbarga University, Gulbarga-585 106, India*

SUMMARY

Pethidine at the dose level of 0.5 mg and 0.75 mg/100 g body weight administered for 20 days to the cycling albino rats caused decrease in the ovarian weight and its protein content. The ovarian folliculogenesis in treated rats is hampered; as a result the follicles which are at the different stages of growth underwent regression. Therefore, the number of healthy follicles is reduced and atretic follicles increased. The elevated levels of ovarian cholesterol and decreased level of glycogen in the pethidine treated rats indicates the inhibition brought in steroidogenesis, which is dependent on pituitary gonadotrophins.

Key words: Pethidine; Rats; Healthy follicles; Atretic follicles; Ovarian steroids

INTRODUCTION

Pethidine (meperidine, Demerol, or phenylpiperidine) is a synthetic analgesic drug introduced by Eisleb and Schaumann in 1939. Like other opioids pethidine binds to opioid receptors and exerts its chief pharmacological actions on the CNS. Opioids act on the hypothalamus and inhibit the release of gonadotrophin releasing hormone (GnRH) and corticotropin releasing factor (CRF), thus decreasing the circulating concentrations of luteinising hormone (LH), follicle stimulating hormone (FSH), adrenocorticotrophic hormone (ACTH) and β -endorphin (Jaffe and Martin, 1985). Secretions of pituitary gonadotrophins are regulated by brain and neurons situated in the anterior parts of the hypothalamus that synthesize the GnRH (Krieger *et al.*, 1982;

Teresawa and Davis, 1983). According to several investigators CNS influencing drugs inhibit the release of FSH and LH from the pituitary acting through hypothalamus, blocking the neural stimulus to the gonadotrophin releasing hormone (Blake *et al.*, 1972; Blake, 1974, 1978; Anderson *et al.*, 1982).

Morphine, an opioid administered during parturition inhibits the uterine contractility, disturbs the normal maternal behaviour and reduces the survival of pups. It also affects the established maternal behaviour in lactating rats (Russell and Spears, 1984; Russell *et al.*, 1989). Pethidine is an analgesic used much more commonly than morphine in obstetric practice. It also has opiate actions, probably mediated by μ -opioid receptors thereby having similar effects like that of morphine on inhibition of oxytocin secretion and parturition in rats (Russell *et al.*, 1991). However morphine an opioid has a well established tonic inhibitory influence on the

*Correspondence: Vijaykumar B Malashetty, Department of Biology, Sharnbasveshwar Independent P.U. College of Science, Gulbarga-585 102, India.
E-mail: viju_yes@rediffmail.com

hypothalamo-hypophyseal axis that disturbs the normal reproductive activities (Bruni *et al.*, 1977; Cicero *et al.*, 1977; Johnsten *et al.*, 1982; Ching, 1983; Cicero *et al.*, 1985). Reports on the effects of pethidine on reproduction are scanty. Therefore the present study is aimed to understand the effect of pethidine on ovarian activities which are dependent on hypophyseal gonadotrophins in albino rats.

MATERIALS AND METHODS

Sexually matured, healthy, colony bred virgin male and female rats of Wistar strain, aged 3 months and weighing 150 - 200 g were used for the experiments. The rats were housed in polypropylene cages measuring 12" × 10" × 8", under well ventilated animal house conditions (temperature: 28 - 31°C, photoperiod: 12 h natural light and 12 h dark; humidity: 50 - 55%). The rats were fed with balanced diet as per CFTRI formula and water *ad libitum*. The rats were divided into three groups of six animals each.

Group I : Received 0.2 ml saline/100 g body weight for 30 days.

Group II : Received 0.5 mg pethidine in 0.2 ml saline/100 g body weight for 30 days.

Group III : Received 0.75 mg pethidine in 0.2 ml saline/100 g body weight for 30 days

The treatment was started from estrous phase only, as the ovarian and uterine activities change markedly from one phase to another phase of estrous cycle. The saline or pethidine was administered intraperitoneally everyday between 10:00 and 11:00 AM.

All the rats were sacrificed on 31st day, 24 h after the last treatment. The ovaries were dissected out immediately and separated out from the adherent tissue and weighed to the nearest mg on an electronic balance. Organ from one side of each animal was fixed in Bouin's fluid, embedded in paraffin wax, sectioned at 5 µm, stained with haematoxylin-eosin for histological studies. Follicular

diameter and morphologies were used to classify follicles by using established methods (Hirshfield, 1983; Sanjay and Joshi, 1997) as

Class I: Small preantral follicles (SPAF) (< 90 µm)

Class II: Large preantral follicles (LPAF) (91 - 260 µm)

Class III: Small antral follicles (SAF) (261 - 350 µm)

Class IV: Medium sized antral follicles (MSAF) (351 - 430 µm)

Class V: Large sized antral follicles (LSAF) (431 - 490 µm)

Class VI: Graafian follicles (GF) (> 491 µm)

Follicles under regression were classified depending on the degree of regression as:

Stage IA: Pyknosis in some granulose cells.

Stage IB: Degenerative changes in the entire granulose layer.

Stage IIA : Oocytes with pyknotic nuclei, blocked meiosis in metaphase I (pseudo maturation) and degenerating cumulus cells.

Stage IIB: Characterized by oocytes floating in the antrum with few pyknotic bodies

Morphometric studies of the ovary were made by using stage and ocular micrometer. Organ from the other side was used for biochemical estimations like cholesterol (Peters and Vanslyke, 1946), protein (Lowry *et al.*, 1951) and glycogen (Carrol *et al.*, 1956).

RESULTS

Changes in body weight

There is no significant change in the body weight, after administration of pethidine.

Ovarian gravimetric changes

Administration of 0.5 mg and 0.75 mg pethidine for 30 days has caused highly significant ($P < 0.001$) decrease in the ovarian weight (Table 1).

Biochemical changes

The pethidine administration has shown inhibitory effect on ovarian activities. Cholesterol, the precursor

Table 1. Effect of Pethidine on ovarian gravimetric and biochemical parameters in albino rats

| Treatment | Ovary mg/100g BW | Cholesterol μg/mg | Protein μg/gm | Glycogen μg/gm |
|-------------------|---------------------|----------------------|------------------|-------------------|
| Saline | 55.95 ± 2.04 | 3.26 ± 0.01 | 6.14 ± 0.04 | 2.12 ± 0.04 |
| 0.5 mg Pethidine | 45.87 ± 2.08** | 4.62 ± 0.03* | 4.18 ± 0.05** | 1.80 ± 0.06* |
| 0.75 mg Pethidine | 42.89 ± 1.09** | 5.30 ± 0.05** | 3.80 ± 0.04** | 1.82 ± 0.03* |

Dose: mg/100 g body weight; Duration: 30 days; M ± S.E. = Arithmetic mean ± Standard error; * $P < 0.01$, ** $P < 0.001$, when compared to saline treated control

for steroid biosynthesis is increased significantly ($P < 0.01$) with 0.5 mg and highly significant ($P < 0.001$) with 0.75 mg pethidine administration. The protein content of ovary is decreased highly significantly ($P < 0.001$) with administration of both the doses. Glycogen, the energy reservoir of female reproductive activities is decreased significantly ($P < 0.01$) with both doses (Table 1).

Ovarian follicular kinetics (Figs. 1-3)

Pethidine administration has caused decreased in the number of healthy follicles and increase in the number of regressing follicles.

The number of SPAF (Class I) and LPAF (Class II) is decreased highly significantly ($P < 0.001$) with both the doses of pethidine. This decrease in the number of SAF, MSAF, LSAF and GF (Class III - Class VI) is significant ($P < 0.01$) with 0.5 mg and highly

significant ($P < 0.001$) with 0.75 mg pethidine administration.

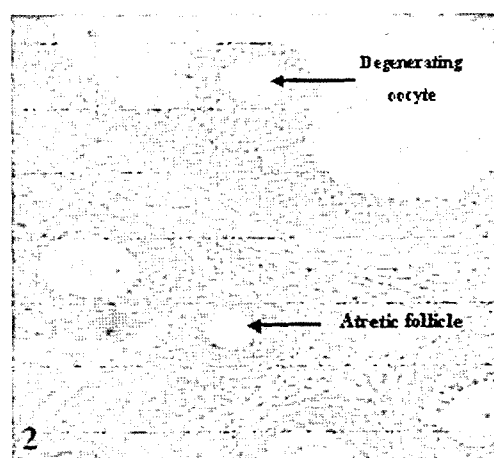


Fig. 2. Treated with 0.5 mg pethidine showing degenerating oocyte and atretic follicles ($\times 400$).

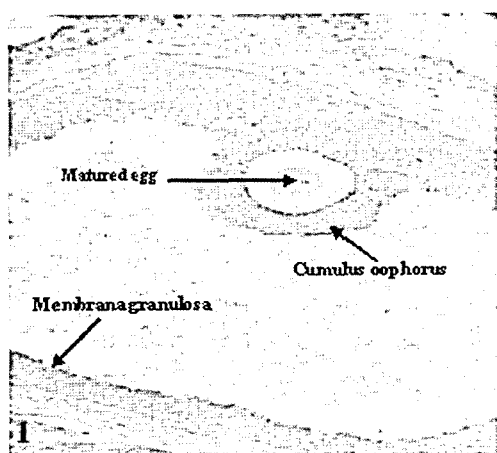


Fig. 1. Control rats with normal ovarian Graafian follicle ($\times 400$).

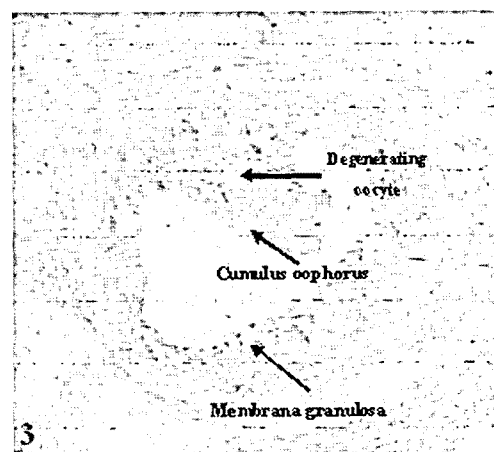


Fig. 3. Treated with 0.75 mg pethidine showing larger follicular atresia with degenerated oocyte, cumulus oophorus and granulosa membrane ($\times 400$).

Table 2. Effect of Pethidine on ovarian follicular kinetics in albino rats

| Treatment | Number of healthy follicles | | | | | |
|-------------------|-------------------------------|--------------------------------------|--------------------------------------|---------------------------------------|--------------------------------------|---------------------------------|
| | Class I < 90 μm | Class II > 91 - 260 μm | Class III > 261-350 μm | Class IV > 351 - 430 μm | Class V > 431 - 490 μm | Class VI > 491 μm |
| Saline | 58.21 \pm 2.81 | 46.67 \pm 0.80 | 11.80 \pm 0.66 | 6.82 \pm 0.42 | 4.08 \pm 0.12 | 2.80 \pm 0.02 |
| 0.5 mg Pethidine | 38.44 \pm 0.2** | 36.26 \pm 1.20** | 7.67 \pm 0.33* | 3.99 \pm 0.29* | 2.72 \pm 0.01* | 1.81 \pm 0.03* |
| 0.75 mg Pethidine | 32.77 \pm 0.91** | 28.23 \pm 1.80** | 4.89 \pm 0.12** | 3.20 \pm 0.24** | 0.94 \pm 0.02** | 0.69 \pm 0.01** |

| Treatment | Number of regressing follicles | | | |
|-------------------|--------------------------------|-------------------|-------------------|-------------------|
| | Stage IA | Stage IB | Stage IIA | Stage IIB |
| Saline | 5.88 \pm 0.04 | 3.47 \pm 0.08 | 1.38 \pm 0.04 | 1.03 \pm 0.01 |
| 0.5 mg Pethidine | 10.29 \pm 0.88** | 7.23 \pm 0.60** | 2.20 \pm 0.09** | 2.08 \pm 0.05** |
| 0.75 mg Pethidine | 13.26 \pm 0.94** | 8.92 \pm 0.24** | 3.33 \pm 0.04** | 3.01 \pm 0.11** |

Dose: mg/100g body weight; Duration: 30 days; M \pm S.E. = Arithmetic mean \pm Standard error; * P < 0.01, ** P < 0.001, when compared to saline treated control.

With both the doses of pethidine administration highly significant (P < 0.001) increase in the number of regressing follicles from stage IA to IIB is observed (Table 2).

DISCUSSION

It is well known that the recruitment of preantral follicles from primary follicles requires pituitary FSH (Young *et al.*, 1992). Further the growth of follicles from preantral stage upto ovulatory stage depends on both FSH and LH. The entire process of follicular growth and cell differentiation appears to involve progressive changes in the response of theca and granulosa cells to LH and FSH respectively (Richards, 1980). The ovarian cycle is a consequence of the dynamic interplay between gonadotrophic stimulation of the ovary and negative and positive feedback actions of ovarian steroids on the hypothalamus and pituitary. Ovulation is triggered through the positive feedback response to estrogen and progesterone involving a midcycle surge of LH (Landgren *et al.*, 1977; Fink, 1979; Crowley and Mac Arthur, 1980).

In mammals endogenous opiod peptides are known to inhibit the release of LH-RH and this action is reversible by its antagonist naloxone (Kalra and Kalra, 1983; Meites, 1986). According to

several investigators CNS influencing drugs inhibit the release of pituitary gonadotrophins from pituitary by acting through hypothalamus thereby blocking the neural stimulus to GnRH (Blake *et al.*, 1972; Blake, 1974, 1978; Anderson *et al.*, 1982). Pethidine, a synthetic opiod analgesic having similar actions of pituitary gonadotrophins via hypothalamo hypophyseal portal system (Eisenman *et al.*, 1958).

In the present study the chronic treatment of pethidine for 30 days has caused the reduction in the weight of ovary. The decrease in the number of follicles from SPAF (Class I) up to ovulating follicles or GF (Class VI) and increase in the number of regressing follicles attributes the non availability of pituitary gonadotrophins due to pethidine treatment as pethidine being a CNS influencing drug and synthetic opiod, inhibits the release of pituitary gonadotrophins (Blake *et al.*, 1972; Blake, 1974, 1978; Anderson *et al.*, 1982). As recruitment of SPAF from primary follicles depends on availability of FSH and further folliculogenesis from SPAF to GF requires FSH and LH (Richards, 1980; Young *et al.*, 1992), more follicles in pethidine treated rats undergo regression rather than maturation.

The low protein content of ovary indicates the retarded ovarian growth as FSH is essential for

protein synthesis in gonads (Means, 1975). High accumulation of cholesterol content in the ovary of drug treated rats may be attributed to the lowered steroidogenesis which is dependent on availability of pituitary gonadotrophins.

The energy source for female reproductive activities is ovarian glycogen, that is estrogen dependent (Walaas, 1952). The supply of glycogen to different reproductive organs in female has been reported to be controlled by the ovarian estrogen and progesterone (Gregoire *et al.*, 1967). The decreased levels of glycogen in the pethidine treated ovary might be due to low ovarian steroidogenesis, which is again attributed to low availability of pituitary gonadotrophins.

REFERENCES

- Anderson K, Fuxe K, Eneroth P, Agnati LF. (1982) Involvement of cholinergic nicotine like receptors as modulators of amine turnover in various terminal systems and prolactin, LH, FSH and ICSH secretion in castrated male rats. *Acta Physiol. Scand.* **116**, 41-50.
- Blake CA, Rex JS, Narman RL, Shigeto K, Sawyer CH. (1972) Effect of nicotine on the proestrous ovulatory surge of LH in the rat. *Endocrinology* **91**, 1253-1259.
- Blake CA. (1974) Localization of inhibitory action of ovulation blocking drugs on release of luteinising hormone in ovariectomised rats. *Endocrinology* **95**, 999-1005.
- Blake CA. (1978) Paradoxical effects of drugs acting on the central nervous system on the preovulatory release of pituitary luteinising hormone in proestrous rats. *J. Endocrinol.* **79**, 319-326.
- Bruni JF, Vanvugt D, Marshall S, Meites J. (1977) Effects of naloxone, morphine and methionine-enkephalin on serum prolactin, luteinising hormone, follicle stimulating hormone and growth hormone. *Life Sci.* **21**, 461-466.
- Caroll NV, Langelly RW, Row RH. (1956) Glycogen determination in liver and muscle by use of anthrone reagent. *J. Biol. Chem.* **26**, 583-593.
- Ching M. (1983) Morphine suppresses the proestrous surge on GnRH in pituitary portal plasma of rats. *Endocrinology* **112**, 2209-2214.
- Cicero TJ, Badger TM, Wilcox CE, Bell RD, Meyer ER. (1977) Morphine decreases luteinising hormone by action on the hypothalamic-pituitary axis. *J. Pharmacol. Ther.* **203**, 548-555.
- Cicero TJ, Schmocker PF, Meyer ER, Miller BT. (1985) Luteinising hormone releasing hormone mediates naloxone effects on serum luteinising hormone levels in normal and morphine sensitized male rats. *Life Sci.* **37**, 467-474.
- Crowley WF, Mac Arthur JW. (1980) Stimulation normal menstrual cycle in Kallawans syndrome by pulsatile administration of luteinising hormone releasing hormone. *J. Clin. Endocrinol. Metab.* **51**, 173-175.
- Eisenman AJ, Fraser HF, Sloan J, Isbell H. (1958) Urinary 17-Ketosteroid excretion during a cycle of addiction to morphine. *J. Pharmacol. Exp. Ther.* **124**, 56-63.
- Fink G. (1979) Feedback actions of target hormones on hypothalamus and pituitary with special reference to gonadal steroids. *Ann. Rev. physiol.* **41**, 571-585.
- Gregoire AT, Ramsay H, Adams A. (1967) The effect of various doses of estradiol and glycogen deposition in the rat uterus, cervix and vagina. *J. Reprod. Fert.* **14**, 231-235.
- Hirshfield AN. (1983) Compensatory ovarian hypertrophy in the long term hemicastrate rat: size distribution of growing and atretic follicles. *Biol. Reprod.* **28**, 271-278.
- Jaff JH, Martin WR. (1985) Opioid analgesics and antagonists. In Goodman LS, Rall WR, Murad F eds. *The pharmacological basis of therapeutics.* Mac Millan Publishing Co., 491-531.
- Johnsten JH, Maughan GT, Anderhuo L. (1982) Inhibition of pulsatile luteinising hormone release by morphine micro injected into the mesencephalic dorsal raphe. *Life Sci.* **30**, 1473-1478.
- Kalra SP, Kalra PS. (1983) Neural regulation of luteinising hormone secretion in rat. *Endocrinol. Rev.* **4**, 311-351.
- Krieger DT, Porlow MJ, Gibson MJ, Davies TF, Zimmerman EA, Ferin M, Gibson (1982) Brain grafts reverse hypogonadism of GnRH deficiency. *Nature* **298**, 468-471.
- Landgren BM, Aedo AR, Nunez M, Cekan SZ,

- Diczfalusy E. (1977) Studies on the pattern of circulating steroids in the normal menstrual cycle, 4, periovulatory changes in relation to the LH surge. *Acta Endocrinol.* **84**, 620-632.
- Lowry DH, Rosenbrough NJ, Farr AL, Randall RJ. (1951) Protein measurement with folin phenol reagent. *J. Biol. Chem.* **193**, 265-275.
- Means AR. (1975) Biochemical effects of follicle stimulating hormone on the testis. In: *Hand Physiol. Sec. J. Endocrinol.* **5**, 203-223.
- Meites J. (1986) Effects of opiates on neuroendocrine functions in animals. In: *Opioid modulation of endocrine function*, Delitalia G, Motta M, Serio M. eds, Raven Press, New York, 53-64.
- Peters JP, Vanslyke DD. (1946) *Quantitative Clinical Chemistry; Vol. I*, Williams and Wilkins eds., Baltimore.
- Richards JS. (1980) Maturation of ovarian follicles: actions and interactions of pituitary and ovarian hormone follicular cell differentiation. *Physiol. Rev.* **60**, 51-89.
- Russell JA, Gosden RG, Humphreys EM, Cutting R, Fitz Simons N, Johnston V, Liddle S, Scott S, Stirland JA. (1989) Interruption of parturition in rats by morphine; a result of oxytocin secretion. *J. Endocrinol.* **121**, 521-536.
- Russell JA, Spears N. (1984) Morphine inhibits suckling induced oxytocin secretion in conscious lactating rats, but also disrupts maternal behaviour. *J. Physiol.* **346**, 133-138.
- Russell JA, Leng G, Coombes JE, Crockett SA, Douglas AJ, Murray I, Way S. (1991) Pethidine (Meperidine) inhibition of oxytocin secretion and action in parturition rats. *Am. J. Physiol.* **261**, 358-368.
- Sanjay VS, Joshi BN. (1997) Melatonin and exposure to constant light/ darkness affects ovarian follicular kinetics and estrous cycle in Indian desert gerbil *Meriones hurrianae* (Jordan). *Gen. Comp. Endocrinol.* **108**, 352-357.
- Terasawa E, Davis GA. (1983) The LHRH neural system in female rats; relation to the medial preoptic nucleus. *Endocrinol. Jpn.* **30**, 405-417.
- Walaas O. (1952) Effect of oestrogens on the glycogen contents of the rat uterus. *Acta Endocrinol.* **10**, 175-192.
- Young EL, Baird DT, Hillier SG. (1992) Mediation gonadotrophin stimulated growth and differentiation on human granulosa cells by 3', 5'-monophosphate: One molecule, two genes. *Clin. Endocrinol.* **37**, 51-55.