RESEARCH ARTICLE

Oridonin Suppresses Proliferation of Human Ovarian Cancer Cells via Blockage of mTOR Signaling

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Abstract

   Oridonin, an ent-kaurane diterpenoid compound isolated from the traditional Chinese herb *Rabdosia rubescens*, has shown various pharmacological and physiological effects such as anti-tumor, anti-bacterial, and anti-inflammatory properties. However, the effect of oridonin on human ovarian cancer cell lines has not been determined. In this study, we demonstrated that oridonin inhibited ovarian cancer cell proliferation, migration and invasion in a dose-dependent manner. Furthermore, we showed oridonin inhibited tumor growth of ovarian cancer cells (SKOV3) *in vivo*. We then assessed mechanisms and found that oridonin specifically abrogated the phosphorylation/activation of mTOR signaling. In summary, our results indicate that oridonin is a potential inhibitor of ovarian cancer by blocking the mTOR signaling pathway.

Keywords: Oridonin - ovarian cancer - mTOR signal pathway - suppress proliferation

Introduction

   Ovarian cancer (OvCa) is the deadliest of all gynecologic cancers; in 2008, more women died from OvCa than from all other gynecologic cancers combined. Each year in the United States, approximately 21,200 women are diagnosed with OvCa (2200<45 years of age), and 14,300 die from the disease (470<45 years of age) (Pollack et al., 2009). The standard therapy for advanced epithelial ovarian cancer (EOC) consists of cytoreductive surgery followed by chemotherapy, preferably including paclitaxel combined with cisplatin or carboplatin (Serkies et al., 2011). It is because of the rational use of operation and chemotherapy, that the estimated 5-year relative survival for women diagnosed with OvCa in 2006 was 44.0%, an increase from 36.1% for those diagnosed between 1975 to 1977(Eheman et al., 2006).But drug toxicity and high price greatly limit the use of these drugs.

   A variety of substances, particularly those present in dietary and medicinal plants, are proposed to have an inhibitory effect on OvCa. Oridonin, an ent-kaurane diterpenoid compound (C20H28O6), isolated from the traditional Chinese herb Rabdosia rubescens, has shown various pharmacological and physiological effects, such as anti-tumor, anti-bacterial, and anti-inflammatory properties (Guo et al., 2013; Bu et al., 2014;Wang et al., 2014). However, the effect of oridonin on human OvCa cell lines has not been determined. Therefore, the present study investigated the effect of oridonin on the migration

and invasion of the OvCa cell line and the underlying molecular mechanism involved.

Materials and Methods

Cell lines, cell culture and reagents
   Oridonin was purchased from Sigma-Aldrich (USA). A 10mM solution of oridonin was prepared in sterile water, stored at -20°C and protected from light, and diluted to needed concentrations for studies. Human ovarian cancer cells (SKOV3,OVCAR-3 and A2780) were purchased from the American Type Culture Collection (ATCC, USA), cultured and maintained in Dulbecco's modified Eagle's medium (DMEM; Hyclone, USA), supplemented with 10% heated-inactivated fetal calf serum (FCS), 100 U/mL penicillin, 10μg/mL streptomycin sulfate and then held at 37°C in a humidified 5% CO_2 incubator. Mitomycin C was ordered from Roche (China).

Cell viability assay
   Human ovarian cancer cells ($2*10^4$ cells∕well) were treated without or with different concentrations of oridonin for 48 h. Cell viability was determined by MTT(Sigma-Aldrich, USA) with Bio-Radmicroplate reader.

Flow cytometry analysis
   Human ovarian cancer cells ($1*10^6$) were treated with different concentrations of oridonin for 48 h and then collected and analyzed for cell cycle distribution in

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a FACS flow cytometer (BD Sciences, USA) with cell cycle Kit (BD Pharmingen, USA).

Wound healing assay

Human ovarian cancer cells were cultured into full confluence in 6-well plates and then were incubated with 10μg/mL mitomycin C for 2 hours to inactivate cell proliferation. Cells were washed with PBS and wounds were made by sterile pipette tips. DMEM supplemented with 0.5% FBS was added into wells with or without different concentrations of oridonin. Cell images were taken after 10 hours of incubation with the aforementioned reagents.

Transwell migration assay

The transwell (Corning Incorporated, USA) were coated with 50μl 1:8 diluted Matrigel (Growth factor reduced, BD Biosciences, USA) for 30min in cell incubator. The bottom chambers of transwell were filled with DMEM with 0.5% FBS and the top chambers were seeded with mitomycin C inactivated, 4×10^4 cells/well in 100μL DMEM (0.5% FBS) plus different concentrations of oridonin. After overnight, the cells on the top surface of the membrane (non-migrated cells) were scraped with a cotton swab and the cells migrated onto the bottom sides of the membrane (invasive cells) were fixed with cold 4% paraformaldehyde and stained with crystal Violet. Images were taken using OLYPUS inverted microscope.

Xenograft mouse model

Xenograft mouse model was used to evaluate the effect of oridonin tumor growth as described by Yi et al (Yi et al., 2008). The 5-week-old to 6-week-old severe combined immune deficiency (SCID) male mice (SLAC Laboratory Animal, China) weighing about 20g were divided into four groups with 6 animals for each group: control group, 5-FU group, oridonin high-dose treated group and oridonin low-dose treated group. SKOV3 cells, 2×10^6 cells per mouse, were s.c. injected into the mice. After the tumors had established (about 100 mm3), oridonin dissolved in sterile PBS, was administered by intraperitoneal injection at 5 or 10mg/kg body weight, respectively. 5-FU, dissolved in sterile PBS, was administered by intraperitoneal injectionat 20-mg/kg/two days body. Control animal was only given 0.2 ml sterile water as vehicle control. The mouse body weight and tumor sizes were recorded every two days and the tumor sizes were determined by Vernier caliper measurements and calculated as 1/2 × length × width × height. After 21 days, mice were sacrificed. Animals used in the present study were approved by the Ethics Committee of Shanghai University of Traditional Chinese Medicine where the work was undertaken and that it conforms to the provisions of the Declaration of Helsinki in 1995.

Western blotting

To determine the effects of oridonin on blocking the phosphorylation of mTOR signaling pathway, cells were first starved with 0.1% FBS medium for 12-14 hours. After washout with new fresh culture medium, cells were pretreated with different concentrations of oridonin for

60 minutes before the phosphorylation status of mTOR signaling molecules was determined. The whole cell extracts were prepared by lysing cells with RIPA buffer supplemented with proteinase inhibitors. Antibodies used for different Western blot analyses, are anti-phospho-mTOR (Ser2448), phospho-4EBP1 (Thr37/46) mAb and phospho-P70S6 (Thr389) (Cell Signaling Technology, USA).

Statistical analysis

The data are presented as mean ± SE, and statistical comparisons between groups were performed using T-test. *P* value ≤0.05 was considered significance statistically.

Results

Inhibition of Ovarian cancer cell lines proliferation by oridonin

Oridonin is a diterpenoid purified from Rabdosia rubescens and has a molecular weight of 364.43 g/mol (Figure 1a). We first tested the in vitro anti-proliferative effect of oridonin on Ovarian cancer cell lines SKOV3, OVCAR-3 and A2780 using a standard proliferation assay (MTT). Our results showed that oridonin significantly reduced SKOV3, OVCAR-3 and A2780 cell proliferation at the IC50 of 17.21μM, 13.9μM and 12.1μM, respectively (Figure 1b, c, d). These data suggest that oridonin preferentially be an anti-OvCa reagent. Further, we examine whether oridonin affected cell cycle progression and apoptosis and our results revealed that it induced a
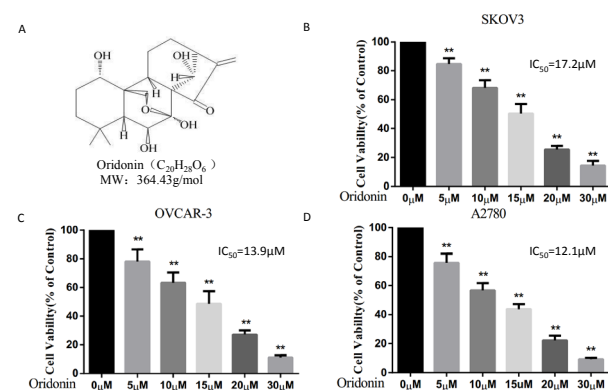

Figure 1. Oridonin Inhibits Cell Viability in OvCa SKOV3, OVCAR-3 and A2780 Cell. (a) The chemical structure of oridonin with a molecular weight 364.43g/mol. (b) Oridonin inhibits SKOV3 cell viability in dose-dependent manner. SKOV3 cell (2*104cells/well) were starved with 0.1% fetal bovine serum (FBS) medium and then treated with or without different concentrations of oridonin for 24 h. Cell viability was quantified by MTT assay (*P< 0.05, **P< 0.01versus Control). (c) Oridonin inhibits OVCAR-3 cell viability in dose-dependent manner. OVCAR-3 cell (2*10⁴ cells/ well) were starved with 0.1% fetal bovine serum (FBS) medium and then treated with or without different concentrations of oridonin for 24 h. Cell viability was quantified by MTT assay (*P< 0.05, **P< 0.01versus Control). (d) Oridonin inhibits A2780 cell viability in dose-dependent manner. A2780 cell (2*10⁴ cells/ well) were starved with 0.1% fetal bovine serum (FBS) medium and then treated with or without different concentrations of oridonin for 24 h. Cell viability was quantified by MTT assay (*P<0.05, **P< 0.01versus Control). Column, mean from three different experiments with six duplicates; bar, SE

G2/M phase block, 44.71% treated vs.19.21% control and a decrease of cells in G1/M phase, from 57.43% to 36.02% (Table 1).
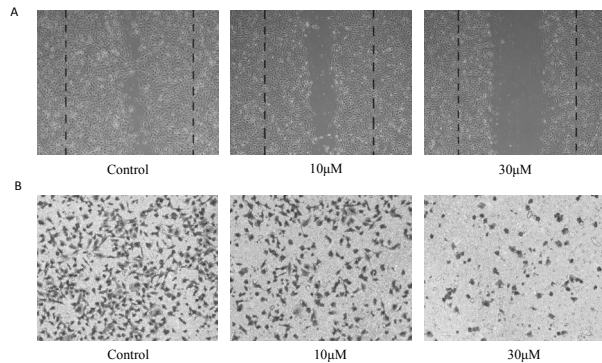

Figure 2. Oridonin Inhibits Migration, Invasion of SKOV3 Cells. (a) Oridonin inhibits SKOV3 cells migration. KOV3 cells were allowed to grow into full confluence in six-well plates, and inactivated with 10lg/mL mitomycin C for 2 h. Cells were wounded with pipette and treated with or without different concentration of oridonin in endothelial cell medium supplemented with 0.5% fetal bovine serum (FBS). After incubation, the migrated cells were quantified by manual counting. (b) Oridonin inhibits the invasion of KOV3 cells. KOV3 cells were seeded in the upper chamber of transwell and treated with different concentrations of oridonin. After about 8-10 h, the invasive KOV3 cells passed through the membrane and were quantified by counting the cells that migrated onto the membrane. Column, mean from three different experiments with duplicates; bar, SE (*P< 0.05, **P< 0.01 versus Control)
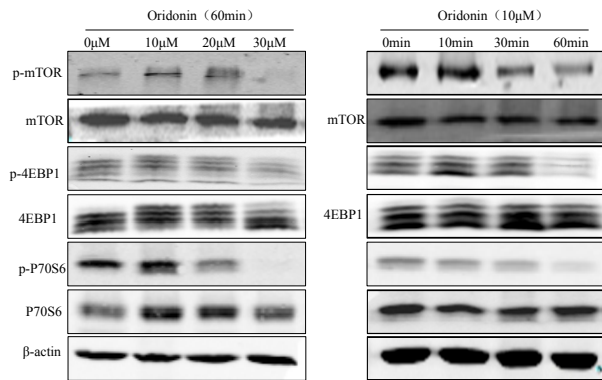

Figure 4. Oridonin Inhibits the mTOR Signaling Pathway in SKOV Cells. (a) Effects of oridonin on blocking the phosphorylation of mTOR signaling pathway. SKOV3 cells were pretreated with various concentrations of oridonin for 60 min. After that, cells were washed with cold phosphate-buffered saline (PBS) and lysed on the dish in RIPA buffer. Phosphorylation and activation of different protein kinases including pSER2248-mTOR, p Thr37/46-4EBP1and pThr389-P70S6 were examined by specific antibodies.(b) A time course of SKOV3 cells without or with the presence of oridonin (30μM): 0,10, 30, 60min

Oridonin inhibits migration, invasion of SKOV3 cells

Tumor migration and invasion are the essential steps in tumorigenesis(Miao et al., 2014). We determined the effects of oridonin on the chemotactic motility of SKOV3 cells using wound-healing migration and transwell cell invasion assays. As shown in Fig2 (a, b), Oridonin significantly inhibited SKOV3 cells migration and invasion at 10μM. Moreover, the inhibition of oridonin on the migration of SKOV3 cells were in a dose-dependent manner.

Oridonin inhibits tumor growth in vivo

We tested oridonin's in vivo effect in a xenograft human ovarian cancer model. Tumor-bearing mice had been treated by oridonin or 5-FU for 21 days. As shown in Fig 3a, tumors in control animals grew rapidly (from 201.59±34.67 to 1494.56±189.77 mm3), whereas tumors in oridonin-treated animals grew much slower (from 132.45±18.47 to 653.12±75.87mm3). Mice treated with oridonin did not have altered body weight (Figure 3b), suggesting no toxicity of the compound at the tested concentration.

Oridonin inhibits the mTOR signaling pathway in SKOV cells

mTOR pathway is a crucial mediator of tumor progression (Zagouri et al., 2012). To examine the
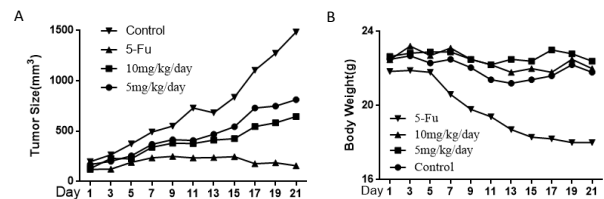

Figure 3. Oridonin Inhibits Tumor Growth *in vivo*. (a) Oridonin inhibits solid cancer growth in xenograft ovarian cancer mouse model. SKOV3 cells were injected (2*10⁶ per mouse) into the 5- to 6-week-old male nude mice. After the tumors had established (about 100 mm3), the mice were injected with or without oridonin at 5 or 10 mg/kg every day, respectively. After 21 days, mice were sacrificed, tumors were removed and photographed. Tumor sizes in control animals and oridonin treated animals were calculated and shown in (a). Tumors in control animals increased from 201.59±34.67 to 1494.56±189.77 mm³, whereas tumors in animals treated with oridonin at dose of 10 mg/kg per day increased from 132.45±18.47 to 653.12±75.87mm³. (b) Treatment with oridonin has little effect on mouse body weight. No significant difference between animals treated with oridonin at dose of 10 mg/kg per day and the control animals. Column, mean; bar, SE (n=7)

Table 1. The Cell Cycle and Apoptosis of SKOV3 after Transfection and Interference by Using Flow Cytometry

Group	n	G0/G1(%)	S(%)	G2/M(%)	Apoptosis(%)
Control	7	57.43±6.23	23.36±3.49	19.21±2.97	1.23±0.21
30uM	7	36.02±5.39**	16.11±1.45	44.71±3.02**	43.47±8.46**

Compared with control P<0.01

underlying molecular mechanism of oridonin's inhibition on proliferation, we studied the effects of oridonin on the phosphorylation of mTOR signaling in SKOV3 cells. Our results showed that oridonin at concentration of 30μM significantly inhibited the phosphorylation of protein kinases involved in the mTOR signaling pathway, including mTOR, 4EBP1, and P70S6 (Figure 4a). We also studied a time course of SKOV3 without or with the presence of oridonin (30μM): 0, 10, 30, 60 min. The results showed that the oridonin got to inhibit the phosphorylation of mTOR signaling at 30 min (Figure 4b).

Discussion

Looking for small molecules which have anti-tumor ability in natural Chinese herbal medicine has been the focus of cancer drug development (Konkimalla et al., 2008;Assaf et al., 2013). In this report, we found that oridonin, an ent-kaurane diterpenoid compound, which has long been used as an anti-inflammation drug, inhibited OvCa tumorigenesis. To further understand the mechanisms in the anti-OvCa activity of oridonin, we analyzed the effect of oridonin on mTOR signaling pathways which were constitutively activated in SKOV3 ovarian cells. It has been shown that oridonin induces apoptosis and attenuates the migration and invasion of melanoma cells and breast cancer cells, as well as decreasing the expression of Bcl-2,caspase-8,NF-κB and PARP proteins(Ren et al., 2006;Wang et al., 2013). Our current studies showed that oridonin repressed SKOV3 cells migration and invasion through inhibiting mTOR signaling pathway. In a xenografts mouse model, we found that oridonin (10mg/kg) did not affect the body weight of the mice, while exhibited significant inhibitory effects on OvCa solid tumor growth.

As we knew, mTOR is directly involved in many cell signaling pathways and many aberrations of the mTOR implicated in human cancer. For example, PI3K amplification/mutation, AKT overexpression, loss of PTEN and P53 function, and overexpression of S6K1 have all been associated with cancer development and are linked to the mTOR signaling pathway(Li et al., 2012;Wang et al., 2014;Xu et al., 2014;Zhang et al., 2014;Zhao et al., 2014). Although mutations of mTOR itself have not been reported, mutations in components of mTOR-related signaling pathways have frequently been described in various human malignancies. Notably, tumor cells with mutations in p53 or PTEN, found in more than 50% of human tumors, suggesting that inhibition of mTOR signaling might be a potential tumor-selective therapeutic strategy(Husseinzadeh et al., 2014). We found that oridonin inhibited the phosphorylation of mTOR in SKOV3 ovarian cells, but did not affect the levels of total mTOR proteins. The reduction of phosphorylated mTOR might explain the observed decrease of the phosphorylation of 4EBP1 and P70S6, the major downstream target of mTOR, following treatment with oridonin. One possible mechanism of how oridonin decreases the phosphorylation of mTOR was that oridonin was directly binding to the kinase domain of mTOR. Our results illustrated that oridonin could significantly inhibit the activation of mTOR signaling

pathway in a concentration-dependent manner with a modest, effective concentration of 30μM.

In summary, this study shows that oridonin inhibits the phosphorylation of mTOR signaling pathway in vitro and tumor tumorigenesis in vivo and is efficacious against the xenograft model of human ovarian cancer in nude mice. Taken together, our studies indicate that oridonin is a potential inhibitor of tumor migration and invasion by blocking mTOR signaling pathway.

Acknowledgements

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